

# Osmotolerant And Osmophilic Fungi From House Dust, Riyadh, Saudi Arabia

H.A. Bokhary

*Dept. of Botany & Microbiology,  
College of Science, King Saud University,  
P.O. Box 2455, Riyadh 11451.  
Saudi Arabia*

**Abstract.** House dust samples collected from filters of vacuum cleaners and air-conditioners filters of different houses in the different localities of Riyadh city were screened for osmotolerant and osmophilic fungi present in the household environment. A total number of 78 fungal species belonging to 31 genera were isolated. Out of these 57 species of 24 genera of fungi were found to grow on M40 (Malt Extract + 40% sucrose) medium (Osmophilic fungi). The total number of fungi per gram of house dust was found to be higher during summer season than in winter. *Aspergillus* was the predominant genus among the 27 species. It was predominant both in summer and winter season and also on M20 and M40 medium, since this genus exhibit more number of species compared to other genera in all the cases. The occurrence of *Aspergillus* was followed by the genera *Eurotium* (7 species) and *Penicillium* (6 species).

**Key Word:** Fungi, Osmotolerant, Osmophilic, House dust, Saudi Arabia.

## Introduction

Modern houses provide a new ecological niche for fungi as a result of reduced ventilation and extensive use of air-conditioning, specially during summer time. Fungi along with mites pose great health hazzards to the residents of these houses (Schoeber, 1988; Rijckaert, 1981; Bokhary and Parvez, 1995). These fungal components, mites and pollen grains cause several allergic diseases like asthma, rhinitis, nasal eosinophilia and dermatitis (Krillis *et al.*, 1985; Chirila *et al.*, 1984; Al-Fryah *et al.*, 1989). Childrens are more susceptible to these diseases than adults (Mori *et al.*, 1985; Bronswijk *et al.*, 1986).

Common molds, usually saprophytic and organisms like *Alternaria*, *Aspergillus*, *Penicillium*, *Mucor*, *Drechslera* and *Cladosporium* found in almost every type of environments are reported to cause allergic asthma (Krillis *et al.* 1985; Al-Doory and Dommson, 1984).

Hamada and Yamada (1991) have recorded *Aspergillus restrictus*, *Eurotium* spp. and *Wellemia sebi* as the most abundant xerophilic (osmophilic) fungal flora of house dust unlike the outdoor airborne fungal flora. Bokhary (1998) has reported *Aspergillus* to be the predominant genus as osmotolerant and osmophilic fungus from sand dunes. Seasonal

fluctuations in the outdoor mycoflora was first studied in Riyadh by Ali *et al.* (1977). Fungi inhabiting household environment in different localities of Riyadh has been reported by Bokhary and Parvez (1995). Fungi were also reported earlier from floor and air-conditioner dust of houses in Riyadh (Saad and El-Gindy 1990). However, no work has so far been reported about osmophilic and osmotolerant fungi from house dust. Therefore, the aim of present study was to investigate the fungi that can survive under extreme condition of low moisture availability or those that are xerophytic in nature (osmotolerant and osmophilic fungi).

### Materials and Methods

Samples of house dust were collected monthly from filters of vacuum cleaners and from filters of air-conditioners in the houses in different localities of Riyadh during the period from January to December 1997. For isolation of fungi, dilution plate method was adopted as earlier (Bokhary and Parvez, 1995). Media used for isolation were M20 (malt extract + 20% sucrose) and M40 (Malt extract + 40% sucrose) for osmotolerant and osmophilic fungi respectively as adopted earlier (Bokhary, 1998). Purification of fungi was carried out every day after 3 days of incubation upto a total of 25 days of incubation. Fungi were identified according to keys cited earlier (Bokhary and Parvez, 1995).

### Results and Discussion

Seasonal fluctuations in the mycoflora of house dust collected from vacuum cleaners are given in Table 1. A total number of 78 fungal species belonging to 31 genera were isolated on M20 (osmotolerant) and M40 (osmophilic) media during different seasons of the year. The highest number of fungal species (71 species) were isolated on M20 medium during May to August period, followed by isolation on M40 medium (55 species) during the same period. The period between September to December yielded second highest numbers of fungi (35 species on M20 medium and 26 species on M40 medium), followed by the period between January to April (29 species on M20 and 23 species on M40 medium). This may be due to the extensive use of air-conditions during the summer (May to August) and also due to less ventilation of houses. *Aspergillus* was the predominant genus and represented by 27 species. *Aspergillus* species were also predominant on M40 medium as most of these could grow on 40% sucrose, hence were able to tolerate high sugar concentrations (osmophilic) or high osmotic potential. Therefore these fungi can survive even at very low humidity condition. Genus *Aspergillus* was followed by genus *Eurotium* with 7 species. *Eurotium* is the ascomycetes phase (perfect state) of *Aspergillus* with visible sclerotia present in the medium. *Penicillium* with 6 species ranked third in dominance. *Penicillium* also could grow well on M40

Table 1. Seasonal fluctuation in the mycoflora of house dust collected from vacuum cleaners.

Fungi	No. of colonies per gram of house dust					
	A		B		C	
	M20	M40	M20	M40	M20	M40
<i>Acremonium strictum</i> W Gams	-	-	12±3	-	-	-
<i>Alternaria alternata</i> Fr.:Fr. Keissler	26±4	20±3	36±5	32±6	25±3	20±4
<i>A. chlamyospora</i> Mouchaca	35±5	26±4	42±6	39±5	20±3	18±3
<i>A. solani</i> Sorauer	-	-	16±3	20±5	-	-
<i>A. tenuissima</i> (Kunze:Pers) Wiltshire	-	-	15±3	-	-	-
<i>Aspergillus candidus</i> Link: Fr.	-	-	23±4	15±3	16±4	-
<i>A. carbonarius</i> (Bain.) Thom	16±3	20±4	39±5	35±4	20±4	16±3
<i>A. carneus</i> (van Tieg.) Blochwitz	-	-	26±4	16±3	-	-
<i>A. chevalieri</i> (Margin) Thom & Church	12±2	-	-	-	-	-
<i>A. conicus</i> Blochwitz	-	-	16±2	12±2	18±3	15±3
<i>A. flavus</i> Link	43±4	36±3	59±7	46±5	49±5	36±5
<i>A. floriformis</i> Samson & Mouchaea	-	-	-	-	11±2	-
<i>A. foetidus</i> Thom & Raper	-	-	-	-	8±3	-
<i>A. fresenii</i> Subram.	8±2	-	9±3	-	12±3	-
<i>A. fumigatus</i> Fresenius	49±8	42±6	58±6	43±5	63±4	49±5
<i>A. japonicus</i> Saito	-	-	20±4	16±3	-	-
<i>A. niger</i> van Tieghem	68±6	57±5	76±8	68±7	63±4	60±5
<i>A. nutans</i> McLennan & Ducker	-	-	12±2	-	-	-
<i>A. niveus</i> Blochwitz	-	-	16±3	10±2	-	-
<i>A. ochraceus</i> Wilhelm	-	-	15±3	12±2	-	-
<i>A. oryzae</i> (Ahlb.) Cohn	12±3	8±2	18±4	12±4	-	-
<i>A. parasiticus</i> Speare	-	-	12±4	-	-	-
<i>A. penicilloides</i> Spegazzini	-	-	18±2	16±3	-	-
<i>A. phoenicis</i> (Corda) Thom	-	-	16±3	-	8±2	-
<i>A. pseudoglaucus</i> Ohtsuki	-	-	8±3	-	6±2	-
<i>A. restrictus</i> G. Smith	8±3	6±2	16±3	12±2	-	-
<i>A. tamarii</i> Kitta	11±3	12±3	19±4	16±4	-	-
<i>A. terreus</i> Thom.	-	-	23±4	16±5	-	-
<i>A. umbrosus</i> Bainier & Sartory	-	-	15±3	15±3	-	-
<i>A. ustus</i> (Bain) Thom et Church	8±3	8±2	18±3	17±3	-	-
<i>A. versicolor</i> (Vuill.) Tiraboschi	-	-	22±4	20±4	16±3	10±2
<i>A. wentii</i> Wehmer	-	-	19±4	24±5	-	-
<i>Aureobasidium pullulans</i> (de Bary) Arnaud	-	-	12±2	15±3	-	-
<i>Bahusaganda</i> sp.	-	-	18±3	16±4	-	-
<i>Basidiobolus haptosporus</i> Drechsler	8±3	-	12±3	-	-	-
<i>Botryotrichum atrogriseum</i> Van Beyma	-	-	18±4	12±3	16±2	14±2
<i>Cephalosporium</i> sp.	-	-	22±3	16±4	-	-
<i>Chaetomium globosum</i>	-	-	16±3	16±4	12±3	10±3
<i>Chalara</i> sp.	-	-	12±3	12±3	-	-
<i>Circinella muscae</i> (Sorok.) Berl. et de Toni	13±2	10±2	18±3	16±2	-	-
<i>Cladosporium cladosporioides</i> (Fres.) De Vries	20±3	18±3	25±5	20±3	29±3	26±4
<i>C. herbarum</i> (Pers: Fr.) Link	-	-	16±3	12±5	-	-
<i>C. sphaerospermum</i> Penzig	-	-	12±3	8±4	19±3	14±3
<i>Cochliobolus spicifer</i> Nelson	-	-	16±3	15±3	-	-
<i>Drechslera</i> sp.	-	-	-	-	8±3	-
<i>Chlamydomyces</i> sp.	13±3	-	18±3	-	-	-
<i>Emericella nidulans</i> (Eidam) Vuill.	-	-	14±3	18±3	8±3	8±2

Table 1 cont.

<i>E. rugulosa</i> (Thom et Raper) Benjamin	-	-	12±3	16±2	-	-	
<i>Embellisia chlamydospora</i> (Hoes & al.) Sin.	-	-	18±3	-	-	-	
<i>Eurotium amstelodami</i> Mangin	14±2	26±4	20±3	32±4	25±4	20±4	
<i>E. chevalieri</i> Mangin	16±3	32±3	29±4	32±4	33±3	39±4	
<i>E. halophilicum</i> Delacr.	13±2	18±3	22±4	31±3	16±2	26±3	
<i>E. montevidensis</i> Talice et Macknnon	-	-	16±3	18±2	-	-	
<i>E. repens</i> de Bary	-	-	-	-	16±4	19±3	
<i>E. rubrum</i> (Konig, spieck., & Brem.) Thom & Church.	20±4	29±4	30±3	42±5	-	-	
<i>E. tonophilum</i> Ohtsuki	-	-	23±4	25±5	-	-	
<i>Fusarium oxysporum</i> Shelecht	24±3	29±4	32±4	39±3	20±3	16±2	
<i>Hyphomyces chrysospermum</i> Tul.	-	-	12±3	14±4	-	-	
<i>Monodictys castaneae</i> (Walter.) Hughes	-	-	11±4	-	-	-	
<i>Mucor hiemalis</i> Wehmer	14±3	-	26±3	-	12±2	-	
<i>Mycelia sterilia</i>	-	-	10±2	12±4	-	-	
<i>Penicillium aurantiogriseum</i> Dierckx	-	-	-	-	14±2	16±3	
<i>P. chrysogenum</i> Thom	36±3	25±4	69±7	63±5	52±3	41±5	
<i>P. citrinum</i> Thom	18±2	10±2	21±3	16±2	-	-	
<i>P. funiculosum</i> Thom	-	-	15±3	14±3	29±3	20±3	
<i>P. oxalicum</i> Currie et Thom	-	-	10±3	12±4	-	-	
<i>P. sclerotiorum</i> Van Beyma	-	-	24±4	-	-	-	
<i>Pleospora herbarum</i> (Fr.) Rabenh. ex les et de Not	-	-	-	-	13±3	12±3	
<i>Rhizopus stolonifer</i> Ehrenb.	18±3	-	29±4	-	16±3	-	
<i>Scopulariopsis brevicaulis</i> (Sacc.) Bainer	11±2	15±3	22±5	20±5	-	-	
<i>S. brumptii</i> Salvanet - Duval	-	-	15±3	12±4	-	-	
<i>Stachybotrys atra</i> Corda	26±4	20±4	39±5	35±5	30±3	25±4	
<i>Torula caligans</i> (Batista & Upadhyay) M.B. Ellis	-	-	10±2	-	-	-	
<i>T. herbarum</i> Link: Fr.	-	-	19±3	12±2	-	-	
<i>Ulocladium atrum</i> Preuss.	18±2	16±2	30±5	30±3	12±3	10±3	
<i>U. chlamydosporum</i> Mouchaca	-	-	16±3	15±3	29±2	26±3	
<i>U. tuberculatum</i> Simmons	-	-	18±2	-	-	-	
<i>Wellemia sebi</i> (Fr.) v. Arx	16±2	18±2	26±3	39±4	22±2	31±4	
Total no. of species	78	29	23	71	55	35	26

- A : Jan. to April  
 B : May to Aug.  
 C : Sept to Dec.  
 ± : Standard deviation.

medium although with lesser number of colonies as compared to its growth on M20 medium. However genus *Eurotium* showed higher number of colonies per gram dust on M40 medium as compared to M20 medium, and therefore could be termed as true osmophilic. *Wellemia sebi* also showed an increased number of colonies per gram of house dust on M40 medium in comparison to M20 medium. Other genera isolated were *Acremonium*, *Alternaria*, *Aureobasidium*, *Bahusaganda*, *Basidiobolus*, *Botryotrichum*, *Cephalosporium*, *Chaetomium*, *Chalara*, *Circinella*, *Cladosporium*, *Cochliobolus*, *Drechslera*, *Chlamydomyces*, *Emericella*, *Embellisia*, *Fusarium*, *Hyphomyces*, *Monodictys*, *Mucor*, *Pleospora*, *Rhizopus*, *Scopulariopsis*, *Stachybotrys*, *Torula* and *Ulocladium*.

The fungal species which were found on both the medium and during all the seasons included *Alternaria alternata*, *A. chlamydospora*, *Aspergillus carbonarius*, *A. flavus*, *A. fumigatus*, *A. niger*, *Cladosporium cladosporioides*, *Eurotium amstelodami*, *E. chevalieri*, *E. halophilicum*, *Fusarium oxysporum*, *Penicillium chrysogenum*, *Stachybotrys atra*, *Ulocladium atrum* and *Wellamia sebi*. Out of 78 fungal species isolated, 57 species were found to grow on M40 medium.

Seasonal fluctuations in the mycoflora of house dust collected from air-conditioners dust are given in Table 2. A total number of 44 fungal species were isolated. The highest number of species were recorded on M20 medium (41 species) during the period May to August followed

by growth on M40 medium (36 species) during the same period. During the period of September to December 24 species were recorded on M20 medium and 21 species on M40 medium. The least number of species were recorded during the period of January to April. This may be due to the lesser use of air-conditioner during this period and more house ventilation as compared to other periods. *Aspergillus* was again the predominant genus exhibiting the highest number of species (12 species) followed by *Eurotium* with six species. *Alternaria alternata*, *A. chlamydospora*, *Aspergillus flavus*, *A. fumigatus*, *A. niger*, *Eurotium halophilicum*, *E. rubrum*, *Penicillium chrysogenum*, *Stachybotrys atra* and *Ulocladium atrum* are the species which were recorded on both media and also during all seasons. Other genera isolated were *Aureobasidium*, *Botryotrichum*, *Cephalosporium*, *Circinella*, *Cladosporium*, *Cochliobolus*, *Emericella*, *Embellisia*, *Fusarium*, *Pleospora*, *Rhizopus*, *Torula*, and *Wellemia*.

*Aspergillus* which yielded the highest number of species and the 3<sup>rd</sup> ranking *Penicillium* were also recorded as the most prevalent genera of house-hold environment and air-conditioner dust of Riyadh (Bokhary and Parvez, 1995; Baggy and Gohar, 1988). However the 2<sup>nd</sup> ranking genus *Eurotium* was not recorded earlier by the same authors. This may be due to the fact that they used normal media for fungal isolation and these *Eurotium* and *Wellemia* species are usually missed because of slow

Table 2. Seasonal fluctuation in the mycoflora of house dust collected from air-conditioner dust.

Fungi	No. of colonies per gram of house dust					
	A		B		C	
	M20	M40	M20	M40	M20	M40
<i>Alternaria alternata</i> Fr. :Fr. Keissler	13±3	12±2	42±5	35±5	20±4	18±3
<i>A. chlamyospora</i> Mouchaca	16±3	15±3	49±4	45±3	33±3	25±4
<i>Aspergillus candidus</i> Link: Fr.	-	-	32±4	14±2	-	-
<i>A. carbonarius</i> (Bain.) Thom	-	-	36±3	32±4	-	-
<i>A. flavus</i> Link.	36±5	30±5	69±7	60±6	42±3	36±4
<i>A. fumigatus</i> Fresenius	33±5	26±4	58±5	52±5	69±3	68±5
<i>A. niger</i> Van Tieghem	48±3	45±5	76±7	70±5	56±4	42±6
<i>A. ochraceus</i> Wilhelm	-	-	28±4	25±5	-	-
<i>A. oryzae</i> (Ahlb.) Cohn	-	-	18±3	16±3	10±2	8±3
<i>A. penicilloides</i> Spegazzini	-	-	29±4	25±5	16±3	12±4
<i>A. restrictus</i> G. Smith	6±2	5±2	25±4	21±3	-	-
<i>A. tamaritii</i> Kitta	-	-	26±4	25±4	-	-
<i>A. terreus</i> Thom.	-	-	16±2	15±3	12±2	10±2
<i>A. ustus</i> (Bain) Thom et Church.	-	-	16±3	16±4	-	-
<i>Aureobasidium pullulans</i> (de Bary) Arnaud	-	-	18±3	16±3	-	-
<i>Botryotrichum atrogriseum</i> Van Beyma	-	-	16±2	15±2	10±2	10±2
<i>Cephalosporium</i> sp.	-	-	26±5	22±-	-	-
<i>Circinella muscae</i> (Sorok.) Berl. et de Toni	-	-	16±2	14±3	-	-
<i>Cladosporium cladosporioides</i> (Fres.) De Varies	40	-	29±4	26±4	16±3	14±3
<i>C. sphaerospermum</i> Penzig	-	-	10±2	6±2	-	-
<i>Cochliobolus spicifer</i> Nelson	-	-	18±3	12±2	-	-
<i>Emericella nidulans</i> (Eidam) Vuill.	-	-	16±2	14±2	-	-
<i>Embellisia chlamyospora</i> (Hoes & al.) Sin	-	-	16±3	-	-	-
<i>Eurotium amstelodami</i> Mangin	-	-	26±4	34±3	15±3	26±4
<i>E. chevalieri</i> Mangin	-	-	36±5	40±5	22±3	26±3
<i>E. halophilicum</i> Dalacr.	10±2	8±2	34±3	35±4	18±2	16±2
<i>E. repens</i> de Bary.	-	-	-	-	10±2	12±3
<i>E. rubrum</i> (Konig, Spieck, & Brem.) Thom & Church	12±2	25±3	36±5	46±4	22±3	32±4
<i>E. tonophilum</i> Ohtsuki	-	-	18±3	16±5	-	-
<i>Fusarium oxysporum</i> Shelecht	-	-	26±3	22±4	16±3	15±4
<i>Mucor hiemalis</i> Wehmer	-	-	26±3	-	29±4	-
<i>Mycelia sterilia</i>	-	-	16±3	14±3	-	-
<i>Penicillium aurantiogriseum</i> Dierckx	-	-	-	-	18±2	-
<i>P. chrysogenum</i> Thom	26±4	25±4	73±5	70±5	56±5	50±4
<i>P. funiculosum</i> Thom.	-	-	18±4	16±3	-	-
<i>P. oxalicum</i> Currie et Thom	-	-	18±3	16±4	-	-
<i>Pleospora herbarum</i> (Ar.) Rabenh. ex les et de Not.	-	-	-	-	8±2	8±3
<i>Rhizopus stolonifer</i> Ehrenb.	14±2	-	32±4	-	15±3	-
<i>Stachybotrys atra</i> Corda	16±2	14±2	29±3	26±4	20±2	18±3
<i>Torula caligans</i> (Batista & Upadhyay) M.B. Ellis.	-	-	16±3	-	-	-
<i>Ulocladium atrum</i> Preuss.	20±3	18±3	36±5	32±5	10±2	10±4
<i>U. chlamyosporum</i> Mouchaca	-	-	22±3	16±2	18±3	16±3
<i>Wellemia sebi</i> (Fr.) v. Arx	-	-	39±4	46±5	-	-
Total no. of species	44					
	12	11	41	36	24	21

A : Jan. to April  
 B : May to Aug.  
 C : Sept. to Dec.  
 ± : Standard deviation.

growth on normal media which are covered by fast growing molds (Hamada and Yamada, 1991). Ali *et al.*, (1977) reported *Cladosporium* as a most prevalent genus as out door air-borne mould flora followed by *Aspergillus*, *Fusarium*, and *Hormodendron*. However this order was not found in the indoorenvironment during the present study and earlier studies (Bokhary and Parvez 1995; Baggy and Gohar, 1988). Common mold genera isolated here like *Aspergillus*, *Penicillium*, *Alternaria*, *Cladosporium*, *Fusarium*, *Ulocladium*, *Mucor*, which are prevalent fungi in house dust, were also common in different types of environments and dusts (Palmgren *et al.*, 1983; Robert *et al.*, 1984; Abdel-Hafez and Shroeit, 1985; Hamada and Yamada, 1986). The fungal species reported here on M20 and M40 medium were also earlier recorded to grow on high sugar concentrations (Bokhary, 1998; Abdel-Hafez, 1982) and to grow even at 50% sucrose concentration (Abdel-Hafez *et al.*, 1990; Hamada and Yamada, 1991). Most of the fungal genera found here were also reported from floor dust of schools (Ali-Shatayel and Arda, 1989; Mercantini *et al.*, 1985; Mercantini *et al.*, 1983). These household inhalant were found to induce positive skin test more frequently than other common allergens. House dust mites are associated with house dust allergens and fungi may stimulate the population growth of the house-dust mites (Samson and Lustgraaf, 1978).

## References

- Abdel-Hafez, S.I.I. 1982. Survey of mycoflora of desert soils in Saudi Arabia. *Mycopathologia*. 80, 1: 3-8.
- Abdel-Hafez, S.I.I., Moubasher, A.H. and Barakat, A. 1990. Keratinophilic fungi and other moulds associated with air-dust particles from Egypt. *Folia Microbiol.* 35: 311-325.
- Abdel-Hafez, S.I.I. and Shoreit, A.A.M. 1985. Mycotoxins producing fungi and mycoflora of air-dust from Taif, Saudi Arabia. *Mycopathologia* 92: 65-71.
- Al-Doory, Y. and Domson, J.F. 1984. Mould Allergy. Philadelphia: Lea and Febiger.
- Al-Fryah, A., Hasnain, S.M., Harfi, H.A. 1989. Respiratory allergy and aero-allergens in Saudi Arabia. *J. Allergy Clin. Immunol.* 83: 198-201.
- Ali, Shatyal, M.S. and Archa, H.M. 1989. Isolation of keratinophilic fungi from floor dust in Arab elementary and preparatory schools in the West Bank of Jordan. *Mycopathologia* 104: 106-111.
- Ali, M.I., Abu-Zinada, A.H. and Al-Mashrawi, Z. 1977. Survey of air-borne mould flora at Riyadh, Saudi Arabia. *Proc. Saudi Biol. Soc.* 1: 215-227.
- Baggy, M.M.K. and Gohar, Y.M. 1988. Mycoflora of air-conditioners dust from Riyadh, Saudi Arabia. *J. Basic Microbiol.* 28: 571-577.
- Bokhary, H.A. 1998. Mycoflora of desert sand-dunes of Riyadh region, Saudi Arabia. *J. King Saud Univ.* 10, 1: 15-29.
- Bokhary, H.A. and Parvez, S. 1995. Fungi inhabiting household environment in Riyadh, Saudi Arabia. *Mycopathologia* 130: 79-87.
- Bronswijk, J.E. M.H. George-Grیدهlet, D.S. and Lustgraaf, B.V. 1986. An evolution of

- biological methods in house dust allergen research. *Allerg. Immunol. (Leipz)*, 24: 18-22.
- Chirila, M., Floresca, L., Popescu, M., Capetti, F. and Panait, F. 1984. The frequency of allergens implicated in bronchial asthma in different areas of Romania. *Rev. Roum. Med. Intern. Med.* 22: 141-146.
- Hamada, N., and Yamada, A. 1991. Seasonal changes in the fungal flora of house dust. *Trans. Mycol. Soc. Japan* 32: 45-53.
- Hudson, H.J. 1980. *Fungal Saprophytism*. Edward Arnold (Pub.) Ltd.
- Krillis, S., Baldo, B.A. and Basten, A. 1985. Analysis of allergen-specific Immunoglobulin E response in 341 allergic patients association between allergens and between allergens groups and clinical diagnosis. *Aust. New Zealand J. Med.* 15: 421-426.
- Mercantini, R., Marsella, R., Lambiase, L. and Belardi, M. 1985. Isolation of keratinophilic fungi from floors in Roman kindergarten and secondary schools. *Mycopathologia* 94: 109-115.
- Mereantini, R., Marsella, R., Lambiase, L. and Fulvi F. 1983. Isolation of keratinophilic fungi from floors in Roman primary schools. *Mycopathologia* 82: 115-120.
- Mori, M., Ogami, Y., and Takashashi, T. 1985. Microbial contamination of carpet dust in private houses. *J. Antibact. Antifung. Agents.* 13: 109-117.
- Palmgren, M.S., Lee, L.S., Delucca, A.J. and Ceigler, A. 1983. Preliminary study of mycoflora and mycotoxin in grain-dust from New Oriens area gram elevators. *Amer. Ind. Hyd. Assoc. J.* 44: 485-488.
- Rijekaert, G. 1981. Exposure to fungi of modern houses. *Allergy* 36: 277-278.
- Robert, A.H., David, M.W. William, R.B. and Odette, L.S. 1984. Viable fungi in corn dust. *Appl. Environ. Microbiol.* 48: 84-87.
- Saad, R., R. and El-Gindy, A.A. 1990. Fungi of house dust in Riyadh, Saudi Arabia. *Zentrabl. Microbiol.* 145: 65-68.
- Samson, R.A. and Lustgraaf, B. 1978. *Aspergillus penicillioides* and *Eurotium halophilicum* in association with house-dust mites. *Mycopathologia* 64: 13-16.
- Schoeber, G. 1988. The influence of the water activity of culture media on the isolation of fungi from house dust. *Mycoses* 31: 255-258.

فطريات غبار المنازل المتحملة والمحبة للضغط الازموزية العالية بمدينة  
الرياض بالمملكة العربية السعودية

أ.د. حسن بن عبد الحكيم بخاري  
قسم النبات والأحياء الدقيقة - كلية العلوم - جامعة الملك سعود  
ص ٥٥٥ ب ٢ الرياض ١١٤٥١ المملكة العربية السعودية

تم تجميع عينات غبار من مرشحات مكانس الشفط ومن مرشحات هواء المكيفات من منازل مختلفة بمدينة الرياض تم فحصها للفطريات المتحملة والمحبة للضغط الازموزية العالية وقد كان العدد الكلي لهذه الفطريات المعزولة هو ٧٨ فطراً تنتمي إلى ٣١ جنساً . وقد وجد من بينها ٥٧ نوعاً تابعة لـ ٢٤ جنس تنمو على بيئة أم - ٤٠ ( مستخلص الشعير + ٤٠ % سكروز ) الخاصة بالفطريات المحبة للضغط الازموزية العالية وقد وجد أن العدد الكلي للفطريات لكل جرام من غبار المنازل يكون أعلى أثناء موسم الصيف عنه من موسم الشتاء وان كان الاسبرجيليس هو الجنس السائد من بين ٢٧ نوع - كما كان سائداً في كلا موسمي الصيف والشتاء وكذلك على وسطي أم ٢٠ وأم ٤٠ حيث أظهر جنس اسبرجيليس و جنس إيروشام ( ٧ أنواع ) و جنس بنسليم ( ٦ أنواع ) .